

BIOGRAPHICAL SKETCH

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NAME: **Jorge D. Miranda**

eRA COMMONS USER NAME (credential, e.g., agency login): **JDMIRANDA**

POSITION TITLE: **Professor**

EDUCATION/TRAINING (*Begin with baccalaureate or other initial professional education, such as nursing, include postdoctoral training and residency training if applicable. Add/delete rows as necessary.*)

| INSTITUTION AND LOCATION | DEGREE (if applicable) | Completion Date MM/YYYY | FIELD OF STUDY |
|---|------------------------------|-------------------------------|----------------|
| University of Puerto Rico, Rio Piedras Campus | B.S. | 05/88 | Biology |
| University of Puerto Rico, Rio Piedras Campus | M.S. | 05/90 | Biology |
| Baylor College of Medicine, Houston, TX | Ph.D. | 10/96 | Neuroscience |
| Univ. of Miami School of Medicine-Miami Project | Postdoc | 12/98 | Regeneration |

A. Personal Statement → website: https://md.rcm.upr.edu/physiology/dt_team/dr-jorge-d-miranda/

The research experience and concepts obtained during my graduate and postdoctoral training support my qualification to accomplish the research project presented in this proposal. As a postdoctoral fellow at the Miami Project to Cure Paralysis, I was exposed to the most recent issues related to spinal cord injury (SCI). Under the mentorship of Dr. Scott Whittemore, he introduced me to the topics, research models and controversial issues in the field of SCI. This research center was an excellent place to perform my postdoctoral training because the environment was suitable to ask questions and share information at all the levels (ions, molecules, proteins, cells, systems, behavior). After two years of postdoctoral training, I joined the Physiology Department at UPR-Medical Sciences Campus in 1999 with a lot of energy and enthusiasm to pass to others what I have learned. At the same time, I developed a collaboration with a group at the Kentucky Spinal Cord Injury Research Center in Louisville, KY. This experience gave me the opportunity to expand my knowledge and skills to answer specific questions related to spinal cord trauma. Initially, the main focus of my laboratory was on the role of repulsive molecules expressed after trauma (Eph receptors and ephrin ligands). However, several new projects were developed. For example, the role of purinergic receptors in the formation of the glial scar, the role of Flotilin-2 lipid raft protein in the clustering of repulsive proteins for axonal regeneration, the role of ephexin (a guanine exchange factor, activated by Eph receptors) in generating the cellular events for growth cone collapse after spinal cord injury and more recently, the neuroprotective role of estradiol and Tamoxifen after injury. Therefore, the technical skills that will be used to achieve the objectives of this proposal are routinely used in my laboratory (& published in several peer review journals). Therefore, no difficulties are expected. So far, I trained three postdoctoral fellows, eight PhD graduate students, and over 20 undergraduate students confirming my motivation and leadership to stimulate others to get involved in science. All my graduate students finished their degree with several publications in peer review journals. Also, my publication record demonstrates that I could work in collaboration with others, confirming my abilities to be a team player. Publications have been developed from collaborations with faculty from the Physiology Department (with Dr. Walter Silva, Dr. Annabell Segarra, Dr. Maria Crespo and Dr. Carlos Jimenez) and also with faculty from other institutions in the USA mainland (Dr. Scott Whittemore, Dr. David Magnuson, and Dr. Marino De Leon). The manuscripts mentioned below endorse our abilities to perform the experiments of interest in our laboratory.

- Willson CA, Irizarry-Ramírez M, Gaskins HE, Cruz-Orengo L, Figueroa JD, Whittemore SR and Miranda JD (2002) Upregulation of EphA Receptor Expression in the Injured Adult spinal Cord. Cell Transplantation 11(3): p.229-239. PMID: 12075988

2. Rosas OR, Santiago JM, Torrado AI, Rodriguez AE, Salgado IK, Miranda JD (2014) Inhibition of Src kinase after spinal cord injury resulted in functional locomotor recovery and increased spared tissue. *Neural Regeneration Research*. 9 (24): 2164-2173. PMID: 25657738; PMC4316450
3. Santiago JM, Rosas O, Torrado AI, González MM, Kalyan-Masih PO, and Miranda JD (2009) Molecular, Anatomical, Physiological and Behavioral studies of rats treated with Buprenorphine. *J. Neurotrauma*. 26 (10), 1783-1793. PMID: 19653810; PMC2864459
4. Figueroa JD, Benton R, Willson CA, Velázquez I, Torrado A, Ortiz C, Whittemore SR and Miranda JD (2006) Inhibition of EphA7 Upregulation after spinal cord injury reduces Apoptosis and Promotes Locomotor Recovery. *J. Neurosc. Res.* 84(7), p. 1438-51. PMID: 16983667

B. Positions and Honors

Positions

- 1986-1988 Undergraduate Research Assistant (Cel. & Mol. Biol. Lab.), University of Puerto Rico
- 1988-1989 Teaching Assistant (General Biology Course), University of Puerto Rico
- 1989-1990 Graduate Research Assistant (Cel. & Mol. Biol. Lab.), University of Puerto Rico
- 1990 Microbiology Teacher, Technological College of San Juan
- 1992-1996 Graduate Research Assistant (Biochemistry Depart.), Baylor College of Medicine
- 1996-1998 Senior Research Associate (Miami Project), University of Miami School of Medicine
- 1999-2004 Assistant Professor, Physiology Department, UPR - School of Medicine
- 2004-2011 Associate Professor, Physiology Department, UPR- School of Medicine (Tenure 2006)
- 2012 Professor, Physiology Department, UPR- School of Medicine
- 2010-2013 Associate Dean and Director of Biomedical Sciences Graduate Program, UPR-Sch. of Medicine
- 2001-2004 Coordinator of the Graduate Program in the Physiology Department
- 2006-2010 Coordinator of the Graduate Program in the Associate Deanship of Biomedical Sciences
- 2006 Vice-President of the Puerto Rico Neuroscience Chapter
- 2007 President of the Puerto Rico Neuroscience Chapter
- 2008-2010 Coordinator of the Human Physiology Course for Medical Students (UPR-Medical Campus)
- 2010-2012 Coordinator of the Human Physiology Course for Dental Students (UPR-Medical Campus)
- 2011 Vice-President of the Puerto Rico Physiological Society Chapter
- 2012 President of the Puerto Rico Physiological Society Chapter
- 2012-2015 Coordinator of the Vertebrate Physiology Course for Graduate Students (UPR-Medical Campus)
- 2017-2023 Co-coordinator of the Human Physiology Course for Medical Students

Honors and Professional Memberships

- 1990-1995 MARC Predoctoral Fellowship (NIGMS)
- 1992-1996 Dean's Award for Excellence, Baylor College of Medicine
- 1995-1996 American Psychological Association Fellowship
- 1996- Member, Society for Neuroscience
- 1996-1997 Research Supplement for Minorities (NIH-NINDS: #NS26887)
- 1997-1998 F32 Postdoctoral Fellowship (NIH-NINDS: #NS10304)
- 1999-2009 Organizing Committee, Puerto Rico Annual Neuroscience Meeting
- 2000-2003 Member, International Society for Developmental Neuroscience
- 2000- Member, Society for Neurotrauma
- 2002 Distinguished Faculty, UPR - School of Medicine (Betances Award)
- 2003-2006 Outstanding Professor, Physiology Department, UPR - School of Medicine
- 2006 Alumni Achievement Award for Research, American Psychological Association (Atlanta,GA)
- 2008-2015 Outstanding Professor, Physiology Department, UPR - School of Medicine
- 2008-2014 Member, American Physiological Society (APS)
- 2009- Member, Puerto Rico Physiological Society (PRPS)
- 2012 Distinguished Faculty, UPR-School of Medicine – Class of Medicine 2012 and 2013
- 2012 Distinguished Faculty, UPR-School of Medicine – Class of Graduate Students 2012
- 2015- Reviewer: *Neural Regeneration Research*
Adhoc Reviewer: 1) *Neurological Research*; 2) *Neuroscience*; 3) *Brain Research*; 4) *Molecular and Cellular Endocrinology*; 5) *Journal of Neurotrauma*; 6) *J. Neurochemistry*
- 2019 & 21 Outstanding Professor, Physiology Department, UPR - School of Medicine

C. Contribution to Science

1. Spinal cord injury triggers a cascade of molecular and cellular events that generates a non-favorable environment for axonal outgrowth and cell survival. Therefore, the need to develop a multi-active drug or agent that tackle many of the events associated with the physical trauma is extremely important to obtain functional locomotor recovery. Initially, we studied the neuroprotective effects of estradiol after SCI in ovariectomized rats. However, long-term treatment with estradiol may produce some adverse effects that may overcome the neuroprotective effects that may create. Therefore, we analyzed the effect of a selective estrogen receptor modulator (Tamoxifen) that activates estrogen receptors without the adverse effects of estradiol. The rationale of the proposed project is to consider Tamoxifen as a neuroprotective agent that could produce a permissive environment for cell survival and axonal elongation, by either transcriptional or non-genomic pathways (like ROS scavengers). We have been able to demonstrate that Tamoxifen confers neuroprotection to the injured spinal cord in a sex-specific manner and with a particular therapeutic window (being longer in females than in male animals). Therefore, with the published data from my laboratory (and others) we need to identify the receptors and intracellular mechanisms activated by Tamoxifen to confers neuroprotection and how this drug affect specifically glial cells at the injury site. Moreover, since we observed a reduction in mechanical allodynia with Tamoxifen, there is a need to recognize the mechanisms activated by this drug to reduce pain after spinal cord injury. The publications produced, as a PI, in this project are:

- a) Colón JM, González PA, Cajigas A, Maldonado WI, Torrado AI, Santiago JM, Salgado IK and Miranda JD (2018) Continuous Tamoxifen delivery improves locomotor recovery 6 hours after spinal cord injury by neuronal and glial mechanisms in male rats. *Experimental Neurology*. 299: 109-121 PMID: 29037533; PMC5723542
- b) Colón JM, Torrado AI, Cajigas A, Santiago JM, Salgado IK, Arroyo Y and Miranda JD (2016) Tamoxifen administration immediately or 24 hours after spinal cord injury improves locomotor recovery and reduces secondary damage in female rats. *J. of Neurotrauma*. 33(18): 1696-708. PMID: 26896212; PMC5035917
- c) Mosquera L, Colón JM, Santiago JM, Torrado AI, Melendez M, Segarra AC, Rodriguez-Orengo, JF, Miranda JD (2014) Tamoxifen and estradiol improved locomotor function and increased spared tissue in rats after spinal cord injury: their antioxidant effect and role of estrogen receptor alpha. *Brain Research*. 1561: 11-22. PMID: 24637260; PMC4046634
- d) Colon JM and Miranda JD (2016) Tamoxifen: an FDA approved drug with neuroprotective effects for spinal cord injury recovery. *Neural Regeneration Research*. 11(8): 1208-1211. PMID: 27651756; PMC5020807

2. The studies performed in the laboratory during my early career were in the area of Eph receptors after spinal cord injury (SCI). The rationale was that trauma to the spinal cord triggers the expression of Eph receptors and ephrin ligands at the injury site, generating a repulsive environment for axonal regeneration and cell survival. We look at the spatio-temporal profile of several Eph receptors and ligands, at the mRNA and protein level. Moreover, when the expression of EphA7 was blocked with antisense technology after SCI we observed an improvement in locomotor recovery and return of nerve conduction. Other study demonstrated that blockaded of the EphA4 produced an increase in mechanical nociception. In addition, we observed an increase in the expression and activation of ephexin, an intracellular mediator of Eph receptor activation, after SCI. Blockade of the ephexin activation produced an increase in locomotor recovery and white matter spared tissue. The publication of my research project was the first article about Eph receptors after spinal cord injury, and for several years we were the group with most publications of this topic in the field. These findings support the idea that Eph receptors should be considered repellent signals that are activated after SCI and studies in this area should contemplate that some repulsive proteins are necessary to avoid sprouting of nociceptive fibers. I served as the primary investigator in most publications in this area. Some publications are:

- a) Miranda JD, White LA, Willson CA, Marcillo A, Jaggid J and Whittemore SR. (1999) Induction of Eph B3 after spinal cord injury. *Exp.Neurol*. 156, p.218. PMID: 10192794
- b) Irizarry-Ramírez M, Willson CA, Cruz L, Figueroa JD, Velázquez I, Jones H, Foster R, Whittemore SR and Miranda JD (2005) Upregulation of EphA3 Receptors After Spinal Cord Injry. *J. of Neurotrauma* 22(8), p.929-935. PMID: 16083359

- c) Cruz-Orengo L, Velázquez I, Torrado A, Ortiz C, Hernández C, Puig A, Segarra A, Whittemore SR and Miranda JD (2006) Blocking EphA4 upregulation after spinal cord injury results in enhanced chronic pain. *Experimental Neurology*. 202, p.421-433. PMID: 16959251
- d) Rosas O, Figueroa JD, Torrado A, Rivera M, König-Toro F and Miranda JD (2011) Expression and activation of Ephexin Expression is altered after spinal cord injury. *Developmental Neurobiology*. 71(7): 595-607. PMID: 20949525; PMC3514508

3. In addition to the studies mentioned above, we also analyzed the role of purinergic receptors in the astrogliotic response after SCI. Specifically, we analyzed the spatio-temporal expression of P2Y₂ after trauma and the role that this protein has in the development of the gliotic scar. The rationale behind this project was that release of ATP after injury activates purinergic receptors in astrocytes, which initiates the gliotic response and this event may generate a non-permissive environment for axonal regeneration. We observed that blockade of the P2Y₂ activation reduced the gliotic response but also induced an increase in the cavity size. These results support the concept that the scar formation by reactive astrocytes is necessary to encapsulate the cyst formed after the injury. To improve locomotor recovery the expression of repellent molecules by reactive astrocytes should be blocked but not the gliotic response that surrounds the debris, dead cells and the fluid filled cavity. As PI of this project, some publications in this area were:

- a) Rodríguez-Zayas A, Torrado A, Miranda JD (2010) P2Y₂ Receptor Expression is Altered in Rats after Spinal Cord Injury. *International J. of Devel. Neurosci. Int. Journal of Developmental Neurosci.* 28(6), 413-21. PMID: 20619335; PMC3225399
- b) Rodríguez-Zayas AE, Torrado AI, Rosas OR, Santiago JM, Figueroa JD and Miranda JD (2011) Blockade of P2 Nucleotide Receptors After Spinal Cord Injury Reduced the Gliotic Response and Spared Tissue. *J. Mol. Neurosci.* 46(1), 167-176. PMID: 21647706; PMC3522077

4. Most membrane receptors that are activated after SCI and block axonal regeneration are located in lipid rafts. The rationale of this project was to analyze the spatio-temporal profile of Flotillin-2, a lipid raft protein that clusters receptors in domains of the plasma membrane, after SCI and determine the role that this protein has in the non-permissive environment for locomotor recovery. In addition, we investigated the expression pattern of caveolin proteins during differentiation of an astrocytoma cell line.

- a) Martínez NA, Ayala AM, Martínez M, Martínez-Rivera FJ, Miranda JD and Silva WI (2016) Caveolin-1 Regulates the P2Y₂ Receptor Signaling in Human 1321N1 Astrocytoma Cells. *J. Biol. Chem.* 291(3), p.12208-22. PMID: 27129210; PMC4933270
- b) Santiago JM, Torrado AI, Arocho LC, Rosas OR, Rodríguez AE, Toro FK, Salgado IK, Torres YA, Silva WI, Miranda JD. (2013) Expression Profile of Flotillin-2 and Its Pathophysiological Role After Spinal Cord Injury. *J. Mol. Neurosci.* 49(2): 347-59. PMID: 22878913; PMC3545048
- c) Silva WI, Maldonado HM, Velázquez G, Rubio-Dávila M, Miranda JD, Aquino E, Mayol N, Cruz-Torres A and Salgado-Villanueva IK (2005) Caveolin isoforms expression during differentiation of C6 glioma cells. *Internat. J. of Developmental Neuroscience* 23, p. 599-612. PMID: 16135403
- d) Martínez M, Martínez N, Miranda JD, Maldonado HM and Silva WI (2019) Caveolin-1 Regulates P2Y₂ Receptor Signaling During Mechanical Injury in Human 1321N1 Astrocytoma. *Biomolecules*. Oct. 18 9(10). PMID: 31635212

Complete List of Published Work in MyBibliography:

<https://www.ncbi.nlm.nih.gov/sites/myncbi/1xKTwoumz7mA8/bibliography/47946842/public/?sort=date&direction=ascending>

D. Additional Information: Research Support and/or Scholastic Performance

Active

| | | |
|-----------|------------------------------|-------------------------------------|
| SC1 | Jorge D. Miranda (PI) | 04/01/2022-03/31/2026 |
| NIH/NIGMS | Role: Principal Investigator | 30% Effort (Total Direct Cost:\$1M) |

The major goal of this grant is to study the effect of Tamoxifen after spinal cord injury in the contractile properties of single muscle fibers in skeletal muscles. Also to evaluate the effect of Tamoxifen on myogenic factors and proliferative proteins from skeletal muscles after spinal cord injury.

COBRE/NIEF José Lasalde (PI) 07/01/23-06/30/28
NIH/NIGMS Role: Director of NIEF 25% Effort (Total NIEF Cost: \$4.5M)

The major goal of this activity is to develop microscopy and electrophysiological core facilities for the use of investigators in the University of Puerto Rico and other institutions in the island, as well as pharmaceutical companies. Technical and state-of-the-art equipment will be available for those users interested to look at high resolution cellular level and functional activity of ion channels in oocytes, cultured neurons, brain slices or anesthetized animals.

NIH/NIMH- MBRS/RISE (T32 G-RISE) Carmen Cadilla (PI) 06/01/23 – 05/31/28
Role: Coordinator Activity #2 4% Effort (Total Direct Cost: \$75K)

Interdepartmental Seminar Series (Activity #2)

The major goal of this activity is to coordinate a monthly seminar series and an annual departmental mini-symposium. This involves the invitation of well-known investigators from the US mainland (different fields of research), and the coordination of seminars (and round table discussions) with the invited speaker and students/faculty.

OVERLAP: There is no scientific overlap between any research proposal and the T32 G-RISE grant.

Completed Support

PR Research, Science & Technology Trust Miranda (PI) 11/2021-09/2022

The major goal of this proposal is to identify the mechanisms used by Tamoxifen to improve locomotor recovery after spinal cord injury in male and female rats.

NIH/NIMH- MBRS/RISE Cadilla (PI) 09/01/04 – 08/31/22

Role: Coordinator Activity #2

The major goal of this activity was to coordinate a monthly seminar series. This involves the invitation of well-known investigators from the US mainland (different fields of research), and the coordination of seminars (and round table discussions) with the invited speaker and students/faculty.

NIH COBRE (1P20GM103642) Miller (PI) 07/01/13 – 12/31/16

Subproject 1: Estradiol and Tamoxifen as neuroprotective/neuroregenerative agents after spinal cord injury

Role: PI of Subproject #1

The major goal of this activity was to determine the neuroprotective and neuroregenerative role of the selective estrogen receptor modulator, tamoxifen, after spinal cord (SCI). Behavioral, anatomical, cellular and molecular analyses were performed to analyze the effect of these agents in the injured spinal cord and establish the therapeutic window of effectiveness.

R24-MH048190 07/01/05 – 06/30/09

NIH/NIMH

M-RISP (Expression Profile and role of Purinergic Receptors After spinal cord injury)

Role: PI of subproject #2

S06-GM08224 08/01/04-07/31/08

NIH/NIGMS

MBRS/SCORE (Role of Eph Receptors during Regeneration of the Nervous system)

Role: PI of subproject #9

U54NS39405-03 09/01/99-08/31/04

NIH/NINDS

The major goal of this subproject was to analyze the spatio-temporal expression of Eph A receptors at the mRNA and protein level after spinal cord injury (SCI) in adult rats.

Role: PI of subproject 2